

Submitted by: Vincent A. Miller, MD
Company: Foundation Medicine, Inc.
Address: 150 Second Street, Cambridge, MA 02141
Phone: 617-418-2259
Email: vmiller@foundationmedicine.com
Date of request: October 25, 2017
NCCN Guidelines Panel: Ovarian Cancer

Dear Panel Members,

On behalf of Foundation Medicine, I respectfully request the NCCN® Ovarian Cancer Guidelines Panel review the enclosed data, which supports comprehensive genomic profiling (CGP) of a tumor specimen as part of the standard of care management for patients with advanced ovarian cancer. CGP assays include FoundationOne®, currently available, and FoundationOne CDx™, currently under parallel and expedited review by FDA and CMS with anticipated approval later this year. We hope that the data accompanying this letter will encourage the NCCN Ovarian Cancer Panel to consider including in its guidelines a recommendation favoring CGP for assessing molecular biomarkers to guide therapy.

Specific Changes: We request that the Panel consider adding CGP to the NCCN Ovarian Cancer Guidelines as part of the work-up for patients experiencing relapse or recurrent disease. CGP is an effective molecular diagnostic tool for the evaluation of a patient with advanced ovarian cancer to identify predictive genomic alterations, including alterations across all exons of *BRCA1* and *BRCA2*, other genes in the homologous recombination repair (HR) pathway, and a phenotype of HR-deficiency (HRD, high levels of genomic loss of heterozygosity; LOH) for potential sensitivity¹⁻³ or resistance^{4,5} to the use of poly (ADP-ribose) polymerase (PARP) inhibitors. CGP also provides a simultaneous assessment of microsatellite instability (MSI), mismatch repair gene alterations that underlie Lynch Syndrome (*MLH1*, *MLH2*, *MLH6*, *PMS2*), and tumor mutational burden (TMB) to guide therapy using immune checkpoint inhibitors such as pembrolizumab⁴. We request that the panel add guidance for the applicability of CGP to support and enhance⁵ the identification of patients with ovarian cancer meeting the NCCN® guidelines for Genetic/Familial High-Risk Assessment⁶. Finally, we request that the panel indicate that CGP may facilitate more rational selection of clinical trials, when appropriate, as it can identify targets for treatment using agents FDA-approved for other indications (e.g. as in the ASCO TAPUR study^{7,8}) or can detect known driver alterations that are rare in a given tumor type and not routinely tested for with disease specific panels, but which confer eligibility for genomically matched clinical trials (e.g. NCI MATCH).

FDA Clearance: FoundationOne is a laboratory developed test (LDT) currently available for clinical use. FoundationOne CDx™ is currently under parallel and expedited review by FDA and CMS with anticipated FDA approval in late 2017. Unlike conventional bridging studies for a single biomarker in one tumor type, achieving an anticipated broad approval involved submitting an analysis across all four classes of genomic alterations (base substitutions, indels, copy number variations and rearrangements) for a dataset comprising more than 6,000 samples. Validation and concordance was demonstrated for more than 36 distinct tumor types and a variety of specimen types (e.g., tumor resections, core biopsies, and fine needle aspirates). The FoundationOne CDx™ assay will both identify patients whose tumors contain alterations associated with FDA-approved therapies and serve as a molecular screen to facilitate access to clinical trials, permitting more rapid testing overall and reducing the time and cost of drug development. This anticipated FDA approved product includes variant calling across 324 genes, genomic signatures for MSI and TMB, and clinical claims in the intended use for diseases with current companion diagnostics, including breast cancer, NSCLC, melanoma, colorectal and ovarian cancers. As such, we continue to submit analogous requests to respective NCCN disease panels for these additional cancers. It is anticipated that this FDA approval across solid tumors will be accompanied by a CMS NCD (National Coverage Determination).

Rationale for Preferring Comprehensive Genomic Profiling: Advanced, metastatic ovarian cancer remains an incurable disease with minimal improvement in mortality over the past decade; prevention within inherited ovarian cancer syndrome families and development of improved treatments are high priorities⁹. Many genes have been shown to be important for the development of ovarian tumors. The use of a broad genomic assay able to detect both specific clinically relevant alterations (e.g., in HR or MMR genes) and complex genomic signatures (i.e., MSI-high or LOH-high) is needed to effectively characterize and categorize ovarian tumors¹⁰. Inherited alterations in several genes predispose women to ovarian cancer, and somatic alterations in many of the same genes also underlie sensitivity to targeted treatments, chemotherapies, and immunotherapies. CGP of a tumor sample using hybrid capture can simultaneously detect both inherited and somatic alterations in *BRCA1*, *BRCA2*, other HR deficiency-related genes¹¹ such as *RAD51C*, *ATM*, or *PALB2*, as well as the

genomic signature of HRD³. Any of these can underlie or predict sensitivity to treatment with inhibitors of PARP¹⁻³ and prompt a genetic/familial high-risk assessment if one had not previously been completed.

A recent analysis shows that only a small fraction of patients with ovarian cancer have undergone ($\leq 11.6\%$) or discussed the NCCN recommended testing (15.1%) for inherited alterations, despite evidence-based guideline support as the standard of care for at least the past 5 years⁵. In addition to helping select effective therapies, CGP may be one method to improve identification of individuals at highest risk for carrying inherited predisposition alleles amongst the estimated 400,000 currently unscreened ovarian cancer patients⁶. Similarly, a fraction of ovarian cancers are related to Lynch Syndrome^{12,13}. Alterations in MMR genes and the phenotypic effects, measured as high levels of MSI and TMB, are detectable using only a sufficiently expansive genomic assay⁴, and may identify both individuals eligible for immunotherapy and additional families with inherited cancer predispositions. Furthermore, unbiased sequencing approaches, such as hybrid capture-based next-generation sequencing, can detect combinations of inherited and acquired genomic alterations (including rearrangements, copy number loss, and insertions or deletions) in HR genes, which may be associated with, depending on their co-occurrence, either sensitivity or resistance to platinum-based chemotherapy or PARP inhibitors¹⁴⁻¹⁷.

For patients with ovarian subtypes or signatures consistent with decreased sensitivity to PARPi, alterations in signaling pathways may prompt exploration of alternative treatment approaches within clinical trials, including combinations, such as olaparib and cediranib¹⁸, or therapies targeting pathways less commonly activated. mTOR/PI3K pathway alterations are found in ~41% of *BRCA*-wildtype/LOH-low (HR proficient) ovarian serous carcinomas¹⁹, 70% of clear cell ovarian cancers⁷, and at high rates in other non-serous subtypes¹⁰. Responses to genomically matched therapies targeting the mTOR/PI3K pathway have been reported, particularly for clear cell carcinomas^{7,8}. The RAS/RAF/MEK pathway is also frequently mutated, especially in low grade serous carcinomas, and may indicate sensitivity to MEK or RAF inhibitors⁸. In addition, several genes are of prognostic significance in ovarian cancer, including *TP53*, *CCNE1*, *KRAS*, *BRAF*, and *NF1*²⁰.

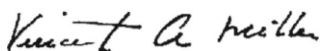
Numerous promising therapeutic approaches are based upon an understanding of cancer genomics and therefore many clinical trials require specified genomic alterations for patient enrollment, including trials offered by the NCI (NCI-MATCH) and ASCO (TAPUR). Consistent with the NCCN[®] recommendation to provide patients with opportunities to participate in clinical trials, multiplex CGP assays, such as FoundationOne[®] and FoundationOne CDx[™], can potentially match > 80% of patients with ovarian cancer to single agent or combination clinical trials. CGP can direct to trials for PARPi and immune modulating therapies¹⁹ by detecting DNA repair pathway alterations¹¹ (affecting *BRCA1*, *BRCA2*, *PALB2*, *BRIP1*, *BARD1*, *ATM*, *POLE*, *POLD1*, *MLH1*, *MSH2*, *MSH6*, and *PMS2*) or complex genomic signatures¹⁹ (LOH-high, MSI-high, or TMB-high). Similarly, mechanistically driven clinical trials may be indicated when alterations affecting other genes: *PIK3CA*, *PTEN*, and *AKT*⁸ (mutation or amplification); *ERBB2* (*HER2*), *ERBB3*, *EGFR*, *FGFR1*, and *FGFR2* (amplification or mutation); *TP53*²¹; *RAS*, *RAF*, or *NF1* alterations are detected^{9,10,22}. Foundation Medicine has joined both the NCI-MATCH and ASCO TAPUR studies as an approved testing platform, and is accelerating accrual to these transformative trials using the combination of CGP and clinical trial matching capabilities. Studies evaluating the use of molecular profiles to direct treatment strategies for patient with gynecological malignancies have shown responses or clinical benefit for 64% of patients²², and CGP has been successfully used to direct patients to clinical trials^{23,24}.

CGP can also inform treatment by refining tumor histology through identification of common, distinguishing, or pathognomonic alterations, such as *FOXL2* (C134W) which distinguishes adult granulosa cell tumors from other sex-cord stromal tumors, or *SMARCA4* alterations that characterize small cell carcinoma of the ovary, hypercalcemic type²⁵ and predict sensitivity to EZH2 inhibitors in clinical development²⁶.

Taken together, these data indicate CGP is an essential addition to clinical care for patients with this deadly malignancy.

Thank you for your review of this submission.

Sincerely,



Vincent A. Miller, M.D.
Chief Medical Officer
Foundation Medicine

References

1. Oza AM, Tinker AV, Oaknin A, et al. Antitumor activity and safety of the PARP inhibitor rucaparib in patients with high-grade ovarian carcinoma and a germline or somatic BRCA1 or BRCA2 mutation: Integrated analysis of data from Study 10 and ARIEL2. *Gynecol Oncol*. September 2017. doi:10.1016/j.ygyno.2017.08.022.
2. Swisher EM, Lin KK, Oza AM, et al. Rucaparib in relapsed, platinum-sensitive high-grade ovarian carcinoma (ARIEL2 Part 1): an international, multicentre, open-label, phase 2 trial. *Lancet Oncol*. 2017;18(1):75-87. doi:10.1016/S1470-2045(16)30559-9.
3. Coleman RL, Oza AM, Lorusso D, et al. Rucaparib maintenance treatment for recurrent ovarian carcinoma after response to platinum therapy (ARIEL3): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Lond Engl*. September 2017. doi:10.1016/S0140-6736(17)32440-6.
4. Chalmers ZR, Connelly CF, Fabrizio D, et al. Analysis of 100,000 human cancer genomes reveals the landscape of tumor mutational burden. *Genome Med*. 2017;9(1):34. doi:10.1186/s13073-017-0424-2.
5. Childers CP, Childers KK, Maggard-Gibbons M, Macinko J. National Estimates of Genetic Testing in Women With a History of Breast or Ovarian Cancer. *J Clin Oncol Off J Am Soc Clin Oncol*. August 2017;JCO2017736314. doi:10.1200/JCO.2017.73.6314.
6. NCCN Clinical Practice Guidelines in Oncology Genetic/Familial High-Risk Assessment: Breast and Ovarian Version 1.2018. October 2017.
7. Elvin JA, Chura J, Gay LM, Markman M. Comprehensive genomic profiling (CGP) of ovarian clear cell carcinomas (OCCC) identifies clinically relevant genomic alterations (CRGA) and targeted therapy options. *Gynecol Oncol Rep*. 2017;20:62-66. doi:10.1016/j.gore.2017.02.007.
8. Castro MP, Whitcomb BP, Zajchowski DA, Coleman RL. Successful use of next generation genomic sequencing (NGS)-directed therapy of clear cell carcinoma of the ovary (CCCO) with trametinib and metformin in a patient with chemotherapy-refractory disease. *Gynecol Oncol Res Pract*. 2015;2. doi:10.1186/s40661-015-0013-2.
9. Matulonis UA, Sood AK, Fallowfield L, Howitt BE, Sehouli J, Karlan BY. Ovarian cancer. *Nat Rev Dis Primer*. 2016;2:16061. doi:10.1038/nrdp.2016.61.
10. Rojas V, Hirshfield KM, Ganesan S, Rodriguez-Rodriguez L. Molecular Characterization of Epithelial Ovarian Cancer: Implications for Diagnosis and Treatment. *Int J Mol Sci*. 2016;17(12):2113. doi:10.3390/ijms17122113.
11. Pennington KP, Walsh T, Harrell MI, et al. Germline and Somatic Mutations in Homologous Recombination Genes Predict Platinum Response and Survival in Ovarian, Fallopian Tube, and Peritoneal Carcinomas. *Clin Cancer Res*. 2014;20(3):764-775. doi:10.1158/1078-0432.CCR-13-2287.
12. Ryan NAJ, Evans DG, Green K, Crosbie EJ. Pathological features and clinical behavior of Lynch syndrome-associated ovarian cancer. *Gynecol Oncol*. 2017;144(3):491-495. doi:10.1016/j.ygyno.2017.01.005.
13. Helder-Woolderink JM, Blok EA, Vasen HFA, Hollema H, Mourits MJ, De Bock GH. Ovarian cancer in Lynch syndrome; a systematic review. *Eur J Cancer Oxf Engl 1990*. 2016;55:65-73. doi:10.1016/j.ejca.2015.12.005.
14. Lord CJ, Ashworth A. BRCAness revisited. *Nat Rev Cancer*. 2016;16(2):110-120. doi:10.1038/nrc.2015.21.
15. Kondrashova O, Nguyen M, Shield-Artin K, et al. Secondary Somatic Mutations Restoring RAD51C and RAD51D Associated with Acquired Resistance to the PARP Inhibitor Rucaparib in High-Grade Ovarian Carcinoma. *Cancer Discov*. 2017;7(9):984-998. doi:10.1158/2159-8290.CD-17-0419.

16. Stover EH, Konstantinopoulos PA, Matulonis UA, Swisher EM. Biomarkers of Response and Resistance to DNA Repair Targeted Therapies. *Clin Cancer Res.* 2016;22(23):5651-5660. doi:10.1158/1078-0432.CCR-16-0247.
17. Norquist B, Wurz KA, Pennil CC, et al. Secondary somatic mutations restoring BRCA1/2 predict chemotherapy resistance in hereditary ovarian carcinomas. *J Clin Oncol Off J Am Soc Clin Oncol.* 2011;29(22):3008-3015. doi:10.1200/JCO.2010.34.2980.
18. Liu JF, Barry WT, Birrer M, et al. Combination cediranib and olaparib versus olaparib alone for women with recurrent platinum-sensitive ovarian cancer: a randomised phase 2 study. *Lancet Oncol.* 2014;15(11):1207-1214. doi:10.1016/S1470-2045(14)70391-2.
19. Elvin JA, He Y, Sun J, et al. Comprehensive genomic profiling (CGP) with loss of heterozygosity (LOH) to identify therapeutically relevant subsets of ovarian cancer (OC). *J Clin Oncol.* 2017;35(15_suppl):5512-5512. doi:10.1200/JCO.2017.35.15_suppl.5512.
20. Patch A-M, Christie EL, Etemadmoghadam D, et al. Whole-genome characterization of chemoresistant ovarian cancer. *Nature.* 2015;521(7553):489-494. doi:10.1038/nature14410.
21. Leijen S, van Geel RMJM, Sonke GS, et al. Phase II Study of WEE1 Inhibitor AZD1775 Plus Carboplatin in Patients With TP53-Mutated Ovarian Cancer Refractory or Resistant to First-Line Therapy Within 3 Months. *J Clin Oncol Off J Am Soc Clin Oncol.* 2016;34(36):4354-4361. doi:10.1200/JCO.2016.67.5942.
22. Rodriguez-Rodriguez L, Hirshfield KM, Rojas V, et al. Use of comprehensive genomic profiling to direct point-of-care management of patients with gynecologic cancers. *Gynecol Oncol.* 2016;141(1):2-9. doi:10.1016/j.ygyno.2016.02.021.
23. Penson RT, Sales E, Sullivan L, et al. A SNaPshot of potentially personalized care: Molecular diagnostics in gynecologic cancer. *Gynecol Oncol.* 2016;141(1):108-112. doi:10.1016/j.ygyno.2016.02.032.
24. Dalton WB, Forde PM, Kang H, et al. Personalized Medicine in the Oncology Clinic: Implementation and Outcomes of the Johns Hopkins Molecular Tumor Board. *JCO Precis Oncol.* 2017;(1):1-19. doi:10.1200/PO.16.00046.
25. Karnezis AN, Cho KR, Gilks CB, Pearce CL, Huntsman DG. The disparate origins of ovarian cancers: pathogenesis and prevention strategies. *Nat Rev Cancer.* 2017;17(1):65-74. doi:10.1038/nrc.2016.113.
26. Wang Y, Chen SY, Karnezis AN, et al. The histone methyltransferase EZH2 is a therapeutic target in small cell carcinoma of the ovary, hypercalcaemic type. *J Pathol.* 2017;242(3):371-383. doi:10.1002/path.4912.